Processing of recorded slices for immunochemical identification // Biocytin staining

Fix brain slices in 4% PFA overnight, then transfer them into 30% sucrose in PBS (Slices can stay in the sucrose solution for a few days. Generally the sooner the processing is done, the better. Can add cryoprotective solution to store them in a -20°c freezer)

Before immunostaining of the recorded slices,

- 1. Get a new and clean 24-well tray, and transfer slices to the tray with a paint brush. Label the try and individual wells. Please write legibly.
- 2. Rinse slices with PBS 3-4 times (3')
- 3. Incubate slices in blocker for **1 hour** [blocker solution for 50 ml: Normal donkey serum 5 ml, BSA, 1 g, Triton X 100, 125 ul, 1xPBS, 45 ml] on a shaker. (blocker: 0.4 ml per well)
- 4. Immunochemical staining: staining biocytin inside the recorded cell with 1:1000 or 1:500 streptavidin-cy3 (Cy3-conjugated Streptavidin Code Number: 016-160-084; Jackson ImmunoResearch Laboratories, Inc) // Incubate slices with streptavidin-cy3 in blocker solution for at least 5 hours at room temperature on a shaker. (streptavidin-cy3 blocker solution 0.4 ml per well). Or could leave the slices overnight at -4 fridge after tested trails for your samples
- 5. Rinse slices with PBS 3-4 times (5') to stop the cy3 staining
- 6. DAPI staining for 10 minutes (to make DAPI solution, add 100 ul DAPI stock solution in 10 ml PBS). 0.4 ml per well // Rinse the slices off with PBS 3-4 times
- 7. Wet-mount slices (on non-subbed slides) and cover-slip with Vectshield.
- 8. Fluorescent scope examination, and take low power picts (x4, x10, x20)
- 9. Confocal scanning (x10, x20, x40)

[Optional procedures:

Remove slices, rinse, and transfer them into 30% sucrose PBS overnight

Resection 400 um slices into 80 um sections

Check some re-sectioned slices to see if cy3 label still exists

Take a few superficial sections for CO staining (for identification laser alignment markers). Use the slice with a cell body to do further immuno staining.]